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(54) Title: METHODS AND REAGENTS FOR DECREASING ALLERGIC REACTIONS

(57) Abstract

It has been determined that allergens, which are characterized by both humoral (IgE) and cellular (T cell) binding sites, can be modified to be less allergenic by modifying the IgE binding sites. The IgE binding sites can be converted to non-IgE binding sites by masking the site with a compound that prevents IgE binding or by altering as little as a single amino acid within the protein, most typically a hydrophobic residue towards the center of the IgE-binding epitope, to eliminate IgE binding. The method allows the protein to be altered as minimally as possible, other than within the IgE-binding sites, while retaining the ability of the protein to activate T cells, and, in some embodiments by not significantly altering or decreasing IgG binding capacity. The examples use peanut allergens to demonstrate alteration of IgE binding sites. The critical amino acids within each of the IgE binding epitopes of the peanut protein that are important to immunoglobulin binding have been determined. Substitution of even a single amino acid within each of the epitopes led to loss of IgE binding. Although the epitopes shared no common amino acid sequence motif, the hydrophobic residues located in the center of the epitope appeared to be most critical to IgE binding.

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METHODS AND REAGENTS FOR DECREASING ALLERGIC REACTIONS

Background of the Invention

The United States government has rights in this invention by virtue of grants from the National Institute of Health RO1-AI33596.

Allergic disease is a common health problem affecting humans and companion animals (mainly dogs and cats) alike. Allergies exist to foods, molds, grasses, trees, insects, pets, fleas, ticks and other substances present in the environment. It is estimated that up to 8% of young children and 2% of adults have allergic reactions just to foods alone. Some allergic reactions (especially those to foods and insects) can be so severe as to be life threatening. Problems in animals tend to be less severe, but very common. For example, many dogs and cats have allergies to flea saliva proteins, grasses, and other common substances present in the environment.

Allergy is manifested by the release of histamines and other mediators of inflammation by mast cells which are triggered into action when IgE antibodies bound to their receptors on the mast cell surface are cross linked by antigen. Other than avoidance, and drugs (e.g. antihistamines, decongestants, and steroids) that only treat symptoms and can have unfortunate side effects and often only provide temporary relief, the only currently medically accepted treatment for allergies is immunotherapy. Immunotherapy involves the repeated injection of allergen extracts, over a period of years, to desensitize a patient to the allergen. Unfortunately, traditional immunotherapy is time consuming, usually involving years of treatment, and often fails to achieve its goal of desensitizing the patient to the allergen. Furthermore, it is not the recommended treatment for food allergies, such as peanut allergies, due to the risk of anaphylaxis.

Noon (Noon, *Lancet* 1911; 1:1572-73) first introduced allergen injection immunotherapy in 1911, a practice based primarily on empiricism with non-standardized extracts of variable quality. More recently the introduction of standardized extracts has made it possible to increase the efficacy of immunotherapy, and double-blind placebo-controlled trials have

demonstrated the efficacy of this form of therapy in allergic rhinitis, asthma and bee-sting hypersensitivity (BSAC Working Party, *Clin. Exp. Allergy* 1993; 23:1-44). However, increased risk of anaphylactic reactions has accompanied this increased efficacy. For example, initial trials of immunotherapy to food allergens has demonstrated an unacceptable safety:efficacy ratio (Oppenheimer et al. *J Allergy Clin. Immun.* 1992; 90:256-62; Sampson, *J. Allergy Clin. Immun.* 1992; 90:151-52; Nelson et al. *J. Allergy Clin. Immun.* 1996; 99:744-751). Results like these have prompted investigators to seek alternative forms of immunotherapy as well as to seek other forms of treatment.

Initial trials with allergen-non-specific anti-IgE antibodies to deplete the patient of allergen-specific IgE antibodies have shown early promise (Boulet, et al. 1997; 155:1835-1840; Fahy, et al. *American J Respir. Crit. Care Med.* 1997; 155:1828-1834; Demoly P. and Bousquet J. *American J Resp. Crit. Care Med.* 1997; 155:1825-1827). On the other hand, trials utilizing immunogenic peptides (representing T cell epitopes) have been disappointing (Norman, et al. *J. Aller. Clin. Immunol.* 1997; 99:S127). Another form of allergen-specific immunotherapy which utilizes injection of plasmid DNA (Raz et al. *Proc. Nat. Acad. Sci. USA* 1994; 91:9519-9523; Hz et al. *Int. Immunol.* 1996; 8:1405-1411) remains unproven.

There remains a need for a safe and efficacious therapy for allergies, especially those where traditional immunotherapy is ill advised due to risk to the patient or lack of efficacy. There is also a need for alternatives to therapies, for example, by creating foods, materials or substances that do not include the allergens that are most problematic, or which contain modified allergens which do not elicit the same reaction. While the technology to make genetically engineered plants and animals is at this point well established, useful modifications would require understanding how allergens can be modified so that they retain the essential functions for the plants' and animals' nutritional value, taste characteristics, etc., but no longer elicit as severe an allergic response.

It is therefore an object of the present invention to provide a method for decreasing the allergenicity of allergens either by modifying the allergen

itself or by producing a compound that would mask the epitope and thus prevent binding of IgE.

It is a further object of the present invention to provide allergens that elicit fewer IgE mediated responses.

It is still another object of the present invention to provide a method to make genetically engineered plants and animals that elicit less of an allergic response than the naturally occurring organisms.

Summary of the Invention

It has been determined that allergens, which are characterized by both humoral (IgG and IgE) and cellular (T cell) binding sites, can be made less allergenic by modifying the IgE binding sites. The IgE binding sites can be eliminated by masking the site with a compound that would prevent IgE binding or by altering as little as a single amino acid within the protein to eliminate IgE binding. The method allows the protein to be altered as minimally as possible, (i.e. only within the IgE-binding sites) while retaining the ability of the protein to activate T cells and, optionally, to bind IgG. Binding sites are identified using known techniques, such as by binding with antibodies in pooled sera obtained from individuals known to be immunoreactive with the allergen to be modified. Proteins that are modified to alter IgE binding are screened for binding with IgG and/or activation of T cells.

Peanut allergens (Ara h 1, Ara h 2, and Ara h 3) have been used in the examples to demonstrate alteration of IgE binding sites while retaining binding to IgG and activation of T cells. The critical amino acids within each of the IgE binding epitopes of the peanut protein that are important to immunoglobulin binding were determined. Substitution of even a single amino acid within each of the epitopes led to loss of IgE binding. Although the epitopes shared no common amino acid sequence motif, the hydrophobic residues located in the center of the epitope appeared to be most critical to IgE binding.

Standard techniques such as a skin test for wheal and flare formation can be used to assess decreased allergenicity of modified proteins, created as described in the examples. The modified allergens can also be tested for

binding to IgG and proliferation of T cells, and modified allergens selected for optimal stimulation of T cells and binding IgG.

The immunotherapeutics can be delivered by standard techniques, using injection, by aerosol, sublingually, topically (including to a mucosal surface), and by gene therapy (for example, by injection of the gene encoding the immunotherapeutic into muscle or skin where it is transiently expressed for a time sufficient to induce tolerance).

This method and the criteria for identifying and altering allergens can be used to design useful proteins (including nucleotide molecules encoding the proteins) for use in immunotherapy, to make a vaccine and to genetically engineer organisms such as plants and animals which then produce proteins with less likelihood of eliciting an IgE response. Techniques for engineering plants and animals are well known. Based on the information obtained using the method described in the examples, one can engineer plants or animals to cause either site specific mutations in the gene encoding the protein(s) of interest, or to knock out the gene and then insert the gene encoding the modified protein.

Brief Description of the Drawings

Figure 1 shows an example of how IgE binding epitopes were mapped to a specific amino acid sequence on the Ara h 1 allergen.

Figure 2 shows how IgE binding epitopes were mapped to a specific amino acid sequence on the Ara h 2 allergen.

Figure 3 shows how IgE binding epitopes were mapped to a specific amino acid sequence on the Ara h 3 allergen.

Figure 4 is a graph of the %IgE binding relative to wild type Ara h2 of modified Ara h 2 allergens.

Figure 5 shows the results of T-cell proliferation assays using the native and recombinant wild-type and modified Ara h 2 protein, compared to crude peanut as a control.

Detailed Description of the Invention

Definitions

The following definitions are used herein.

An antigen is a molecule that elicits production of antibody (a

humoral response) or an antigen-specific reaction with T cells (a cellular response).

An allergen is a subset of antigens which elicits IgE production in addition to other isotypes of antibodies.

An allergic reaction is one that is IgE mediated with clinical symptoms primarily involving the cutaneous (uticaria, angiodema, pruritus), respiratory (wheezing, coughing, laryngeal edema, rhinorrhea, watery/itching eyes), gastrointestinal (vomiting, abdominal pain, diarrhea), and cardiovascular (if a systemic reaction occurs) systems.

An epitope is a binding site including an amino acid motif of between approximately six and fifteen amino acids which can be bound by either an immunoglobulin or recognized by a T cell receptor when presented by an antigen presenting cell in conjunction with the major histocompatibility complex (MHC). A linear epitope is one where the amino acids are recognized in the context of a simple linear sequence. A conformational epitope is one where the amino acids are recognized in the context of a particular three dimensional structure.

An immunodominant epitope is one which is bound by antibody in a large percentage of the sensitized population or where the titer of the antibody is high, relative to the percentage or titer of antibody reaction to other epitopes present in the same protein.

A decreased allergic reaction is characterized by a decrease in clinical symptoms following treatment of symptoms associated with exposure to an allergen, which can involve respiratory, gastrointestinal, skin, eyes, ears and mucosal surfaces in general.

An antigen presenting cell (an APC) is a cell which processes and presents peptides to T cells to elicit an antigen-specific response.

Immunostimulatory sequences are oligodeoxynucleotides of bacterial, viral or invertebrate origin that are taken-up by APCs and activate them to express certain membrane receptors (e.g., B7-1 and B7-2) and secrete various cytokines (e.g., IL-1, IL-6, IL-12, TNF). These oligodeoxynucleotides containing unmethylated CpG motifs cause brisk activation and when injected into animals in conjunction with antigen, appear

to skew the immune response to a Th1-type response. See, for example, Yamamoto, et al., *Microbiol. Immunol.* 36, 983 (1992); Krieg, et al., *Nature* 374, 546-548 (1995); Pisetsky, *Immunity* 5, 303 (1996); and Zimmerman, et al., *J. Immunol.* 160, 3627-3630 (1998).

I. Diagnostic and Therapeutic Reagents.

The first step in making the modified allergen is to identify IgE binding sites and/or immunodominant IgE binding sites. The second step is to mutate one or more of the IgE binding sites, preferably including at a minimum one of the immunodominant sites, or to react the allergen with a compound that selectively blocks binding to one or more of the IgE binding sites. The third step is to make sufficient amounts of the modified allergen for administration to persons or animals in need of tolerance to the allergen, where the modified allergen is administered in a dosage and for a time to induce tolerance, or for diagnostic purposes. The modified allergen can be administered by injection, or in some cases, by ingestion or inhalation.

A. Allergens.

Many allergens are known that elicit allergic responses, which may range in severity from mildly irritating to life-threatening. Food allergies are mediated through the interaction of IgE to specific proteins contained within the food. Examples of common food allergens include proteins from peanuts, milk, grains such as wheat and barley, soybeans, eggs, fish, crustaceans, and mollusks. These account for greater than 90% of the food allergies (Taylor, Food Techn. 39, 146-152 (1992). The IgE binding epitopes from the major allergens of cow milk (Ball, et al. (1994) *Clin. Exp. Allergy*, 24, 758-764), egg (Cooke, S.K. and Sampson, H.R. (1997) *J. Immunol.*, 159, 2026-2032), codfish (Aas, K., and Elsayed, S. (1975) *Dev. Biol. Stand.* 29, 90-98), hazel nut (Elsayed, et al. (1989) *Int. Arch. Allergy Appl. Immunol.* 89, 410-415), peanut (Burks et al., (1997) *Eur. J. Biochemistry*, 245:334-339; Stanley et al., (1997) *Archives of Biochemistry and Biophysics*, 342:244-253), soybean (Herein, et al. (1990) *Int. Arch. Allergy Appl. Immunol.* 92, 193-198) and shrimp (Shanty, et al. (1993) *J. Immunol.* 151, 5354-5363) have all been elucidated, as have others. Other allergens include proteins from insects such as flea, tick, mite, fire ant,

cockroach, and bee as well as molds, dust, grasses, trees, weeds, and proteins from mammals including horses, dogs, cats, etc.

The majority of allergens discussed above elicit a reaction when ingested, inhaled, or injected. Allergens can also elicit a reaction based solely on contact with the skin. Latex is a well known example. Latex products are manufactured from a milky fluid derived from the rubber tree, *Hevea brasiliensis* and other processing chemicals. A number of the proteins in latex can cause a range of allergic reactions. Many products contain latex, such as medical supplies and personal protective equipment. Three types of reactions can occur in persons sensitive to latex: irritant contact dermatitis, and immediate systemic hypersensitivity. Additionally, the proteins responsible for the allergic reactions can fasten to the powder of latex gloves. This powder can be inhaled, causing exposure through the lungs. Proteins found in latex that interact with IgE antibodies were characterized by two-dimensional electrophoresis. Protein fractions of 56, 45, 30, 20, 14, and less than 6.5 kd were detected (Posch A. et al., (1997) J. *Allergy Clin. Immunol.* 99(3), 385-395). Acidic proteins in the 8-14 kd and 22 - 24 kd range that reacted with IgE antibodies were also identified (Posch A. et al., *(1997) J. Allergy Clin. Immunol.* 99(3), 385-395. The proteins prohevein and hevein, from hevea brasiliensis, are known to be major latex allergens and to interact with IgE (Alenius, H., et al., *Clin. Exp. Allergy* 25(7), 659-665; Chen Z., et al., (1997) *J. Allergy Clin. Immunol.* 99(3), 402-409). Most of the IgE binding domains have been shown to be in the hevein domain rather than the domain specific for prohevein (Chen Z., et al., (1997) *J. Allergy Clin. Immunol.* 99(3), 402-409). The main IgE-binding epitope of prohevein is thought to be in the N-terminal, 43 amino acid fragment (Alenius H., et al., (1996) *J. Immunol.* 156(4), 1618-1625). The hevein lectin family of proteins has been shown to have homology with potato lectin and snake venom disintegrins (platelet aggregation inhibitors) (Kielisqewski, M.L., et al., (1994) *Plant J.* 5(6), 849-861).

B. Identification of IgE Binding Sites.

Allergens typically have both IgE and IgG binding sites and are recognized by T cells. The binding sites can be determined either by using

phage display libraries to identify conformational epitopes (Eichler and Houghten, (1995) *Molecular Medicine Today* 1, 174-180; Jensen-Jarolim et al., (1997) *J. Appl. Clin. Immunol.* 101, 5153a) or by using defined peptides derived from the known amino acid sequence of an allergen (see examples below), or by binding of whole protein or protein fragments to antibodies, typically antibodies obtained from a pooled patient population known to be allergic to the allergen. It is desirable to modify allergens to diminish binding to IgE while retaining their ability to activate T cells and in some embodiments by not significantly altering or decreasing IgG binding capacity. This requires modification of one or more IgE binding sites in the allergen.

A preferred modified allergen is one that can be used with a majority of patients having a particular allergy. Use of pooled sera from allergic patients allows determination of one or more immunodominant epitopes in the allergen. Once some or all of the IgE binding sites are known, it is possible to modify the gene encoding the allergen, using site directed mutagenesis by any of a number of techniques, to produce a modified allergen as described below, and thereby express modified allergens. It is also possible to react the allergen with a compound that achieves the same result as the selective mutation, by making the IgE binding sites inaccessible, but not preventing the modified allergen from activating T cells, and, in some embodiments, by not significantly altering or decreasing IgG binding.

Assays to assess an immunologic change after the administration of the modified allergen are known to those skilled in the art. Conventional assays include RAST (Sampson and Albergo, 1984), ELISAs (Burks, et al. 1986) immunoblotting (Burks, et al. 1988), and *in vivo* skin tests (Sampson and Albergo 1984). Objective clinical symptoms can be monitored before

and after the administration of the modified allergen to determine any change in the clinical symptoms.

It may be of value to identify IgEs which interact with conformational rather than linear epitopes. Due to the complexity and heterogeneity of patient serum, it may be difficult to employ a standard immobilized allergen affinity-based approach to directly isolate these IgEs in quantities sufficient to permit their characterization. These problems can be avoided by isolating some or all of the IgEs which interact with conformational epitopes from a combinatorial IgE phage display library.

Steinberger et al. (Steinberger, P., Kraft D. and Valenta R. (1996) "Construction of a combinatorial IgE library from an allergic patient: Isolation and characterization of human IgE Fabs with specificity for the major Timothy Grass pollen antigen," Phl p. 5 *J. Biol. Chem.* 271, 10967-10972) prepared a combinatorial IgE phage display library from mRNA isolated from the peripheral blood mononuclear cells of a grass allergic patient. Allergen-specific IgEs were selected by panning filamentous phage expressing IgE Fabs on their surfaces against allergen immobilized on the wells of 96 well microtiter plates. The cDNAs were than isolated from allergen-binding phage and transformed into E coli for the production of large quantities of monoclonal, recombinant, allergen-specific IgE Fabs.

If native allergen or full length recombinant allergen is used in the panning step to isolate phage, then Fabs corresponding to IgEs specific for conformational epitopes should be included among the allergen-specific clones identified. By screening the individual recombinant IgE Fabs against denatured antigen or against the relevant linear epitopes identified for a given antigen, the subset of conformation-specific clones which do not bind to linear epitopes can be defined.

To determine whether the library screening has yielded a complete inventory of the allergen-specific IgEs present in patient serum, an immunocompetition assay can be performed. Pooled recombinant Fabs would be preincubated with immobilized allergen. After washing to remove unbound Fab, the immobilized allergen would then be incubated with patient serum. After washing to remove unbound serum proteins, an incubation

with a reporter-coupled secondary antibody specific for IgE Fc domain would be performed. Detection of bound reporter would allow quantitation of the extent to which serum IgE was prevented from binding to allergen by recombinant Fab. Maximal, uncompeted serum IgE binding would be determined using allergen which had not been preincubated with Fab or had been incubated with nonsense Fab. If IgE binding persists in the face of competition from the complete set of allergen-specific IgE Fab clones, this experiment can be repeated using denatured antigen to determine whether the epitopes not represented among the cloned Fabs are linear or conformational.

Production of Recombinant or Modified Allergens

A modified allergen will typically be made using recombinant techniques. Expression in a procaryotic or eucaryotic host including bacteria, yeast, and baculovirus-insect cell systems are typically used to produce large (mg) quantities of the modified allergen. It is also possible to make the allergen synthetically, if the allergen is not too large, for example, less than about 25-40 amino acids in length.

Production of Transgenic Plants and Animals

Transgenic plants or animals expressing the modified allergens have two purposes. First, they can be used as a source of modified allergen for use in immunotherapy and second, appropriately modified plants or animals can be substituted for the original plant or animal, making immunotherapy unnecessary. Furthermore, it is possible that eating modified peanuts or cod fish, for example, could have either or both of two effects: (1) not imparting an allergic response on their own and (2) conferring protection from the unmodified source by acting as an immunotherapeutic agent for the unmodified source. Methods for engineering of plants and animals are well known and have been for a decade. For example, for plants see Day, *(1996) Crit. Rev. Food Sci. & Nut.* 36(S), 549-567, the teachings of which are incorporated herein. See also Fuchs and Astwood (1996) *Food Tech.* 83-88. Methods for making recombinant animals are also well established. See, for example, Colman, A" Production of therapeutic proteins in the milk of transgenic livestock" (1998) *Biochem. Soc. Symp.* 63, 141-147; Espanion and Niemann, (1996) *DTW Dtxch Tierarztl Wochenschr* 103(8-9), 320-328; and

Colman, *Am. J. Clin. Nutr.* 63(4), 639S-6455S, the teachings of which are incorporated herein. One can also induce site specific changes using homologous recombination and/or triplex forming oligomers. See, for example, Rooney and Moore, (1995) *Proc. Natl. Acad. Sci. USA* 92, 2141-2149; Agrawal, et al., *BioWorld Today*, vol. 9, no. 41, p. 1"Chimeriplasty - Gene Surgery, Not Gene Therapy - Fixes Flawed Genomic Sequences" David N. Leff.

Production and Screening of Compounds blocking IgE Binding Sites

Once the IgE binding sites have been identified, it is also possible to block or limit binding to one or more of these sites by reacting the allergen with a compound that does not prevent the allergen from activating T cells, and in some embodiments does not significantly alter or decrease IgG binding capacity, resulting in a modified allergen similar in functionality to that produced by mutation. There are two principal ways to obtain compounds which block IgE binding sites: combinatorial libraries and combinatorial chemistry.

Identification of Compounds That Mask IgE Binding Sites through Application of Combinatorial Chemistry

In some cases it may be preferable to utilize non-peptide compounds to block binding of IgE to the allergen by masking the IgE binding epitope. This can be accomplished by using molecules that are selected from a complex mixture of random molecules in what has been referred to as "*in vitro* genetics" or combinatorial chemistry (Szostak, *TIBS* 19:89, 1992). In this approach a large pool of random and defined sequences is synthesized and then subjected to a selection and enrichment process. The selection and enrichment process involves the binding of the IgE binding epitopes to a solid support, followed by interaction with the products of various combinatorial libraries. Those molecules which do not bind these molecules at all are removed immediately by elution with a suitable solvent. Those molecules which bind to the epitopes will remain bound to the solid support, whereas, unbound compounds will be removed from the column. Those compounds bound to the column can be removed, for example, by competitive binding. Following removal of these compounds, the

compounds which have bound can be identified, using methodology well known to those of skill in the art, to isolate and characterize those compounds which bind to or interact with IgE binding epitopes. The relative binding affinities of these compounds can be compared and optimum compounds identified using competitive binding studies which are well known to those of skill in the art.

Identification of Compounds That Interact with IgE Binding Sites through Application of Combinatorial Phage Display Libraries

Recombinant, monoclonal Fabs directed against conformational epitopes, identified as described above, can be used as reagents to assist in the definition of the biochemical nature of these epitopes. Cross-linking studies employing derivatized Fabs can be employed to label amino acid residues in the vicinity of the epitopes. Similarly, the Fabs can be used in protease protection studies to identify those domains of the allergen protein which are shielded from protease degradation by pre-binding of a specific Fab. Experiments employing recombinant monoclonal Fabs as reagents to label or protect from labeling should permit at least partial elucidation of the structures of conformational epitopes.

"Humanized" recombinant Fabs should bind to allergens if injected into a patient and thus prevent the binding of these allergens to native IgE. Since the Fabs cannot interact with the Fcε receptor, the binding of the IgE Fabs to allergen would not be expected to elicit mast cell degranulation. Allergen should be neutralized as it is by protective IgGs.

Anti-idiotype antibodies directed against the conformational epitope-specific Fabs should resemble the conformation epitopes themselves. Injection of these anti-idiotype antibodies should induce the production of anti-anti-idiotype IgGs which would recognize, bind to and inactivate the conformational epitopes. The method through which the anti-idiotype antibodies would be produced (i.e. animal immunization, "*in vitro*" immunization or recombinant phage display library) would have to be determined. Similarly, the possibility that the anti-idiotype antibodies (which resemble the conformational epitopes) would be recognized by patient IgEs and induce mast cell degranulation needs to be considered.

II. Diagnostic and Therapeutic Procedures Using Modified Allergens.

It is important to administer the modified allergen to an individual (human or animal) to decrease the clinical symptoms of allergic disease by using a method, dosage, and carrier which are effective. Allergen will typically be administered in an appropriate carrier, such as saline or a phosphate saline buffer. Allergen can be administered by injection subcutaneously, intramuscularly, or intraperitoneally (most humans would be treated by subcutaneous injection), by aerosol, inhaled powder, or by ingestion.

Therapy or desensitization with the modified allergens can be used in combination with other therapies, such as allergen-non-specific anti-IgE antibodies to deplete the patient of allergen-specific IgE antibodies (Boulet, et al. (1997) 155:1835-1840; Fahy, et al. (1997) *American J Respir. Crit. Care Med.* 155:1828-1834; Demoly, P. and Bousquet (1997) *J Am J Resp. Crit. Care Med.* 155:1825-1827), or by the pan specific anti-allergy therapy described in U. S. Serial No. 08/090,375 filed June 4, 1998, by M. Caplan and H. Sosin. Therapy with the modified allergen can also be administered in combination with an adjuvant such as IL 12, IL 16, IL 18, Ifn-ζ.

The nucleotide molecule encoding the modified allergen can also be administered directly to the patient, for example, in a suitable expression vector such as a plasmid, which is injected directly into the muscle or dermis, or through administration of genetically engineered cells.

In general, effective dosages will be in the picogram to milligram range, more typically microgram to milligram. Treatment will typically be between twice/weekly and once a month, continuing for up to three to five years, although this is highly dependent on the individual patient response.

The modified allergen can also be used as a diagnostic to characterize the patient's allergies, using techniques such as those described in the examples.

EXAMPLES

Peanut allergy is one of the most common and serious of the immediate hypersensitivity reactions to foods in terms of persistence and

severity of reaction. Unlike the clinical symptoms of many other food allergies, the reactions to peanuts are rarely outgrown, therefore, most diagnosed children will have the disease for a lifetime (Sampson, H.A., and Burks, A.W. (1996) *Annu. Rev. Nutr.* 16, 161-77; Bock, S.A. (1985) *J. Pediatr.* 107, 676-680). The majority of cases of fatal food-induced anaphylaxis involve ingestion of peanuts (Sampson et al., (1992) *NEJM* 327, 380-384; Kaminogawa, S. (1996) *Biosci. Biotech. Biochem.* 60, 1749-1756). The only effective therapeutic option currently available for the prevention of a peanut hypersensitivity reaction is food avoidance. Unfortunately, for a ubiquitous food such as a peanut, the possibility of an inadvertent ingestion is great.

The examples described below demonstrate identification, modification, and assessment of allergenicity of the major peanut allergens, Ara h 1, Ara h 2, and Ara h 3. Detailed experimental procedures are included for Example 1. These same procedures were used for Examples 2-5. The nucleotide sequences of Ara h 1, Ara h 2, and Ara h 3, are shown in SEQ ID NOs. 1, 3, and 5, respectively. The amino acid sequences of Ara h 1, Ara h 2, and Ara h 3 are shown in SEQ ID NOs. 2, 4, and 6 respectively.

Example 1: Identification of linear IgE binding epitopes.

Due to the significance of the allergic reaction and the widening use of peanuts as protein extenders in processed foods, there is increasing interest in defining the allergenic proteins and exploring ways to decrease the risk to the peanut-sensitive individual. Various studies over the last several years have identified the major allergens in peanuts as belonging to different families of seed storage proteins (Burks, et al. (1997) *Eur. J. Biochem.* 245, 334-339; Stanley, et al. (1997) *Arch. Biochem. Biophys.* 342, 244-253). The major peanut allergens Ara h 1, Ara h 2, and Ara h 3 belong to the vicilin, conglutin and glycinin families of seed storage proteins, respectively. These allergens are abundant proteins found in peanuts and are recognized by serum IgE from greater than 95% of peanut sensitive individuals, indicating that they are the major allergens involved in the clinical etiology of this disease (Burks, et al. (1995) *J. Clinical Invest.*, 96, 1715-1721). The genes encoding Ara h 1 (SEQ ID NO. 1), Ara h 2 (SEQ ID NO. 3), and Ara h 3

(SEQ ID NO. 5) and the proteins encoded by these genes (SEQ ID NO. 2, 4, 6) have been isolated and characterized. The following studies were conducted to identify the IgE epitopes of these allergens recognized by a population of peanut hypersensitive patients and a means for modifying their affinity for IgE.

Experimental Procedures

Serum IgE. Serum from 15 patients with documented peanut hypersensitivity reactions (mean age, 25 yrs) was used to determine relative binding affinities between wild type and mutant Ara h 1 synthesized epitopes. The patients had either a positive double-blind, placebo-controlled, food challenge or a convincing history of peanut anaphylaxis (laryngeal edema, severe wheezing, and/or hypotension; Burks, et al. (1988) *J. Pediatr.* 113, 447-451). At least 5 ml of venous blood was drawn from each patient, allowed to clot, and serum was collected. A serum pool from 12 to 15 patients was made by mixing equal aliquots of serum IgE from each patient. The pools were then used in immunoblot analysis.

Peptide synthesis. Individual peptides were synthesized on a derivatized cellulose membrane using 9-fluorenyllmethoxycarbonyl (Fmoc) amino acid active esters according to the manufacturer's instructions (Genosys Biotechnologies, Woodlands, Texas; Fields, G.B. and Noble, R.L. (1990) *Int. J. Peptide Protein Res.* 35, 161-214). Fmoc-amino acids (N-terminal blocked) with protected side chains were coupled in the presence of 1-methyl-2-pyrrolidone to a derivatized cellulose membrane. Following washing with dimethylformamide (DMF), unreacted terminal amino groups were blocked from further reactions by acetylation with acetic anhydride. The N-terminal Fmoc blocking group was then removed by reaction with 20% piperidine and 80% DMF, v/v. The membrane was washed in DMF followed by methanol, the next reactive Fmoc-amino acid was then coupled as before, and the sequence of reactions was repeated with the next amino acid. When peptide synthesis was complete, the side chains were deprotected with a mixture of dichloromethane (DCM), triflouroacetic acid, and triisobutylsilane (1.0:1.0:0.5), followed by successive washes in DCM, DMF, and methanol. Peptides synthesis reactions were monitored by

bromophenol blue color reactions during certain steps of synthesis. Cellulose derivitised membranes and Fmoc-amino acids were supplied by Genosys Biotechnologies. All other chemical were purchased from Aldrich Chemical Company, Inc. (Milwaukee, WI) or Fluka (Buchs, Switzerland). Membranes were either probed immediately or stored at -20°C until needed.

IgE binding assays. Cellulose membranes containing synthesized peptides were washed 3 times in Tris-buffered saline (TBS; 136 mM NaCl, 2.7 mM KCl, and 50 mM trizma base pH 8.0) for 10 min at room temperature (RT) and then incubated overnight in blocking buffer: [TBS, 0.05% Tween™ 20; concentrated membrane blocking buffer supplied by Genosys; and sucrose (0.0:1.0:0.5)]. The membrane was then incubated in pooled sera diluted in 1:5 in 20 mM Tris-Cl pH7.5, 150 mM NaCl, and 1% bovine serum albumin overnight at 4°C. Primary antibody was detected with ^{125}I-labeled equine anti-human IgE (Kallestad, Chaska, MN).

Quantitation of IgE binding. Relative amounts of IgE binding to individual peptides were determined by a Bio-Rad (Hercules, CA) model GS-700 imaging laser densitometer and quantitated with Bio-Rad molecular analyst software. A background area was scanned and subtracted from the obtained values. Following quantitation, wild type intensities were normalized to a value of one and the mutants were calculated as percentages relative to the wild type.

Synthesis and purification of recombinant Ara h 2 protein. cDNA encoding Ara h 2 was placed in the pET-24b expression vector. The pET-24 expression vector places a 6 x histidine tag at the carboxyl end of the inserted protein. The histidine tag allows the recombinant protein to be purified by affinity purification on a nickel column (HisBind resin). Recombinant Ara h 2 was expressed and purified according to the instructions of the pET system manual. Briefly, expression of the recombinant Ara h 2 was induced in 200 ml cultures of strain BL21(DE3) E. coli with 1 mM IPTG at mid log phase. Cultures were allowed to continue for an additional 3 hours at 36°C. Cells were harvested by centrifugation at 2000 x g for 15 minutes and then lysed in denaturing binding buffer (6 M urea, 5 mM imidazole, 0.5 M NaCl, 20 mM Tris-HCl, pH 7.9). Lysates were cleared by centrifugation at 39,000 x g for

20 minutes followed by filtration though 0.45 micron filters. The cleared lysate was applied to a 10 ml column of HisBind resin, washed with imidazole wash buffer (20 mM imidazole, 6 M urea, 0.5 M NaCl, 20 mM Tris-HCl, pH 7.9). The recombinant Ara h 2 was then released from the column using elution buffer (1 M imidazole, 0.5 M NaCl, 20 mM Tris-HCl, pH 7.9). The elution buffer was replaced with phosphate buffered saline by dialysis. The purification of recombinant Ara h 2 was followed by SDS PAGE and immunoblots. Peanut specific serum IgE was used as a primary antibody.

Skin prick tests. The ability of purified native and recombinant Ara h 2 to elicit the IgE mediated degranulation of mast cells was evaluated using prick skin tests in a peanut allergic individual. An individual meeting the criteria for peanut allergy (convincing history or positive double blind placebo controlled food challenge) and a non-allergic control were selected for the testing. Purified native and recombinant Ara h 2 and whole peanut extract (Greer Laboratories, Lenoir, N.C.) were tested. Twenty microliters of the test solution were applied to the forearm of the volunteer and the skin beneath pricked with a sterile needle. Testing was started at the lowest concentration (less than or equal to 1 mg/ml) and increased ten fold each round to the highest concentration or until a positive reaction was observed. Mean diameters of the wheal and erythema were measured and compared to the negative saline control. A positive reaction was defined as a wheal 3mm larger then the negative control. Histamine was used as the positive control.

Results

Identification of the linear IgE-binding epitopes of Ara h 1, Ara h 2 and Ara h 3 allergens. Epitope mapping was performed on the Ara h 1, Ara h 2 and Ara h 3 allergens by synthesizing each of these proteins in 15 amino acid long overlapping peptides that were offset from each other by 8 amino acids. The peptides were then probed with a pool of serum IgE from 15 patients with documented peanut hypersensitivity. This analysis resulted in multiple IgE binding regions being identified for each allergen. The exact position of each IgE binding epitope was then determined by re-synthesizing these IgE reactive regions as 10 or 15 amino acid long peptides that were

offset from each other by two amino acids. These peptides were probed with the same pool of serum IgE from peanut sensitive patients as used before. An example of this procedure for each of the peanut allergens is shown in Figures 1-3. Figure 1 shows amino acid residues 82-133 of Ara h 1, containing peptides 4, 5, 6, and 7, as identified in Table 1. Figure 2 shows amino acid residues 55-76 of Ara h 2, containing peptides 6 and 7, as shown in Table 2. Figure 3 shows amino acid residues 299-321 of Ara h 3, containing peptide 4 as identified in Table 3. This analysis revealed that there were 23 linear IgE binding epitopes on Ara h 1, 10 epitopes on Ara h 2, and 4 epitopes on Ara h 3.

In an effort to determine which, if any, of the epitopes were recognized by the majority of patients with peanut hypersensitivity, each set of epitopes identified for the peanut allergens were synthesized and then probed individually with serum IgE from 10 different patients. All of the patient sera tested recognized multiple epitopes.

Table 1 shows the amino acid sequence and position of each epitope within the Ara h 1 protein of all 23 IgE binding epitopes mapped to this molecule. Table 2 shows the amino acid sequence and position of each epitope within the Ara h 2 protein of all 10 IgE binding epitopes mapped to this molecule. Table 3 shows the amino acid sequence and position of each epitope within the Ara h 3 protein of all 4 IgE binding epitopes mapped to this molecule.

Four epitopes of the Ara h 1 allergen (peptides 1, 3, 4, 17 of Table 1), three epitopes of the Ara h 2 allergen (peptides 3, 6, 7 of Table 2), and 1 epitope of the Ara h 3 allergen (peptide 2 of Table 3) were immunodominant.

Table 1. Ara h I IgE Binding Epitopes

EPITOPE	AA SEQUENCE	POSITION
1	AKSSPYQKKT	25-34
2	QEPDDLKQKA	48-57
3	LEYDPRLVYD	65-74
4	GERTRGRQPG	89-98
5	PGDYDDDRRQ	97-106
6	PRREEGGRWG	107-116
7	REREEDWRQP	123-132
8	EDWRRPSHQQ	134-143
9	QPRKIRPEGR	143-152
10	TPGQFEDFFP	294-303
11	SYLQEFSRNT	311-320
12	FNAEFNEIRR	325-334
13	EQEERGQRRW	344-353
14	DITNPINLRE	393-402
15	NNFGKLFEVK	409-418
16	GTGNLELVAV	461-470
17	RRYTARLKEG	498-507
18	ELHLLGFGIN	525-534
19	HRIFLAGDKD	539-548
20	IDQIEKQAKD	551-560
21	KDLAFPGSGE	559-568
22	KESHFVSARP	578-587
23	PEKESPEKED	597-606

The underlined portions of each peptide are the smallest IgE binding sequences as determined by this analysis. All of these sequences can be found in SEQ ID NO 2.

Table 2. Ara h 2 IgE Binding Epitopes

EPITOPE	AA SEQUENCE	POSITION
1	HASARQQWEL	15-24
2	QWELQGDRRC	21-30
3	DRRCQSQLER	27-36
4	LRPCEQHLMQ	39-48
5	KIQRDEDSYE	49-58
6	YERDPYSPSQ	57-66
7	SQDPYSPSPY	65-74
8	DRLQGRQQEQ	115-124
9	KRELRNLPQQ	127-136
10	QRCDLDVESG	143-152

The underlined portions of each peptide are the smallest IgE binding sequences as determined by this analysis. All of these sequences can be

found in SEQ ID NO 4.

Table 3. Ara h 3 IgE Binding Epitopes

EPITOPE	AA SEQUENCE	POSITION
1	IETWNPNNQEFECAG	33-47
2	GNIFSGFTPEFLEQA	240-254
3	VTVRGGLRILSPDRK	279-293
4	DEDEYEYDEEDRRRG	303-317

The underlined portions of each peptide are the smallest IgE binding sequences as determined by this analysis. All of these sequences can be found in SEQ ID NO 6.

Example 2 : Modification of peanut allergens to decrease allergenicity.

The major linear IgE binding epitopes of the peanut allergens were mapped using overlapping peptides synthesized on an activated cellulose membrane and pooled serum IgE from 15 peanut sensitive patients, as described in Example 1. The size of the epitopes ranged from six to fifteen amino acids in length. The amino acids essential to IgE binding in each of the epitopes were determined by synthesizing duplicate peptides with single amino acid changes at each position. These peptides were then probed with pooled serum IgE from 15 patients with peanut hypersensitivity to determine if the changes affected peanut-specific IgE binding. For example, epitope 9 in Table 1 was synthesized with an alanine or methionine residue substituted for one of the amino acids and probed. The following amino acids were substituted (first letter is the one-letter amino acid code for the residue normally at the position, the residue number, followed by the amino acid that was substituted for this residue; the numbers indicate the position of each residue in the Ara h 1 protein, SEQ ID NO. 2): Q143A, P144A; R145A; K146A; I147A; R148A; P149A; E150A; G151A; R152A; Q143M; P144M; R145M; K146M; I147M; R148M; P149M; E150M; G151M; R152M. The immunoblot strip containing the wild-type and mutated peptides of epitope 9 showed that binding of pooled serum IgE to individual peptides was dramatically reduced when either alanine or methionine was substituted for each of the amino acids at positions 144, 145, and 147-150 of Ara h 1 shown in SEQ ID NO. 2. Changes at positions 144, 145, 147, and 148 of Ara h 1 shown in SEQ ID NO. 2 had the most dramatic effect when methionine was substituted for the wild-type amino acid, resulting in less than 1% of peanut specific IgE binding to these peptides. In contrast, the substitution of an alanine for arginine at position 152 of Ara h 1 shown in SEQ ID NO. 2 resulted in increased IgE binding. The remaining Ara h 1 epitopes, and the Ara h 2 and Ara h 3 epitopes, were tested in the same manner and the intensity of IgE binding to each spot was determined as a percentage of IgE binding to the wild-type peptide. Any amino acid substitution that resulted in less than 1% of IgE binding when compared to the wild type peptide was noted and is indicated in Tables 4-6. Table 4 shows the amino acids that

were determined to be critical to IgE binding in each of the Ara h 1 epitopes. Table 5 shows the amino acids that were determined to be critical to IgE binding in each of the Ara h 2 epitopes. Table 6 shows the amino acids that were determined to be critical to IgE binding in each of the Ara h 3 epitopes.

This analysis indicated that each epitope could be mutated to a non-IgE binding-peptide by the substitution of a single amino acid residue.

The results discussed above for Ara h 1, Ara h 2, and Ara h 3 demonstrate that once an IgE binding site has been identified, it is possible to reduce IgE binding to this site by altering a single amino acid of the epitope. The observation that alteration of a single amino acid leads to the loss of IgE binding in a population of peanut-sensitive individuals is significant because it suggests that while each patient may display a polyclonal IgE reaction to a particular allergen, IgE from different patients that recognize the same epitope must interact with that epitope in a similar fashion. Besides finding that many epitopes contained more than one residue critical for IgE binding, it was also determined that more than one residue type (ala or met) could be substituted at certain positions in an epitope with similar results. This allows for the design of a hypoallergenic protein that would be effective at blunting allergic reactions for a population of peanut sensitive individuals. Furthermore, the creation of a plant producing a peanut where the IgE binding epitopes of the major allergens have been removed should prevent the development of peanut hypersensitivity in individuals genetically predisposed to this food allergy.

Table 4: **Amino Acids Critical to IgE Binding of Ara h 1**

EPITOPE	AA SEQUENCE	POSITION
1	AKSSPYQKKT	25-34
2	QEPDDLKQKA	48-57
3	LEYDPRLVYD	65-74
4	GERTRGRQPG	89-98
5	PGDYDDDRRQ	97-106
6	PRREEGGRWG	107-116
7	REREEDWRQP	123-132
8	EDWRRPSHQQ	134-143
9	QPRKIRPEGR	143-152
10	TPGQFEDFFP	294-303
11	SYLQEFSRNT	311-320
12	FNAEFNEIRR	325-334
13	EQEERGQRRW	344-353
14	DITNPINLRE	393-402
15	NNFGKLFEVK	409-418
17	RRYTARLKEG	498-507
18	ELHLLGFGIN	525-534
19	HRIFLAGDKD	539-548
20	IDQIEKQAKD	551-560
21	KDLAFPGSGE	559-568
22	KESHFVSARP	578-587

Note. The Ara h 1 IgE binding epitopes are indicated as the single letter amino acid code. The position of each peptide with respect to the Ara h 1 protein is indicated in the right hand column. The amino acids that, when altered, lead to loss of IgE binding are shown as the bold, underlined residues. Epitopes 16 and 23 were not included in this study because they were recognized by a single patient who was no longer available to the study. All of these sequences can be found in SEQ ID NO 2.

Table 5. Amino Acids Critical to IgE Binding of Ara h 2

EPITOPE	AA SEQUENCE	POSITION
1	HASAR**Q**Q**W**EL	15-24
2	Q**W**E**L**Q**G**DRRC	21-30
3	D**RR**C**Q**SQL**ER**	27-36
4	**L**R**P**CE**QH**LMQ	39-48
5	**K**IQ**RD**E**D**SYE	49-58
6	YER**DPY**SPSQ	57-66
7	SQ**DPY**SPSPY	65-74
8	DRL**QGR**QQEQ	115-124
9	**KR**E**L**RN**L**PQQ	127-136
10	QRC**DL**D**VE**SG	143-152

Note. The Ara h 2 IgE binding epitopes are indicated as the single letter amino acid code. The position of each peptide with respect to the Ara h 2 protein is indicated in the right hand column. The amino acids that, when altered, lead to loss of IgE binding are shown as the bold, underlined residues. All of these sequences can be found in SEQ ID NO 4.

Table 6. Amino Acids Critical to IgE-Binding of Ara h 3.

EPITOPE	AA SEQUENCE	POSITION
1	IETWN**PN**NQEFECAG	33-47
2	GN**IF**SG**F**TPE**FL**EQA	240-254
3	VTVRGG**L**R**IL**S**P**DRK	279-293
4	DEDEY**EYDE**E**DR**RRG	303-317

Note. The Ara h 3 IgE binding epitopes are indicated as the single letter amino acid code. The position of each peptide with respect to the Ara h 3 protein is indicated in the right hand column. The amino acids that, when altered, lead to loss of IgE binding are shown as the bold, underlined All of these sequences can be found in SEQ ID NO 6.

Example 3: A Modified Ara h 2 Protein Binds less IgE But Similar Amounts of IgG.

In order to determine the effect of changes to multiple epitopes within the context of the intact allergen, four epitopes (including the three immunodominant epitopes) of the Ara h 2 allergen were mutagenized and the protein produced recombinantly. The amino acids at position 20, 31, 60, and 67 of the Ara h 2 protein (shown in SEQ ID NO. 4) were changed to alanine by mutagenizing the gene encoding this protein by standard techniques. These residues are located in epitopes 1, 3, 6, and 7 and represent amino acids critical to IgE binding that were determined in Example 2. The modified and wild-type versions of this protein were produced and immunoblot analysis performed using serum from peanut sensitive patients. These results showed that the modified version of this allergen bound significantly less IgE than the wild type version of these recombinant proteins (Figure 4) but bound similar amounts of IgG.

Example 4: A modified Ara h 2 protein retains the ability to stimulate T-cells to proliferate.

The modified recombinant Ara h 2 protein described in Example 3 was used in T-cell proliferation assays to determine if it retained the ability to activate T cells from peanut sensitive individuals. Proliferation assays were performed on T-cell lines grown in short-term culture developed from six peanut sensitive patients. T-cells lines were stimulated with either 50 μg of crude peanut extract, 10 μg of native Ara h 2, 10 μg of recombinant wild-type Ara h2, or 10 μg of modified recombinant Ara h 2 protein and the amount of 3H-thymidine determined for each cell line. Results were expressed as the average stimulation index (SI) which reflected the fold increase in 3H-thymidine incorporation exhibited by cells challenged with allergen when compared with media treated controls (Figure 5).

Example 5: A Modified Ara h 2 Protein Elicits a Smaller Wheal and Flare in Skin Prick Tests of a Peanut Sensitive Individual.

The modified recombinant Ara h 2 protein described in Example 3 and the wild type version of this recombinant protein were used in a skin prick test of a peanut sensitive individual. Ten micrograms of these proteins were applied separately to the forearm of a peanut sensitive individual, the skin pricked with a sterile needle, and 10 minutes later any wheal and flare that developed was measured. The wheal and flare produced by the wild-type Ara h 2 protein (8 mm X 7 mm) was approximately twice as large as that produced by the modified Ara h 2 protein (4 mm X 3mm). A control subject (no peanut hypersensitivity) tested with the same proteins had no visible wheal and flare but, as expected, gave positive results when challenged with histamine. In addition, the test subject gave no positive results when tested with PBS alone. These results indicate that an allergen with only 40% of its IgE binding epitopes modified (4/10) can give measurable reduction in reactivity in an *in vivo* test of a peanut sensitive patient.

These same techniques can be used with the other known peanut allergens, Ara h 1 (SEQ ID NO 1 and 2), Ara h 3 (SEQ ID NO. 5 and 6), or any other allergen.

We claim:

1. A method of making a modified allergen which is less reactive with IgE comprising

(a) identifying IgE binding sites in an allergen;

(b) modifying the allergen by mutating at least one amino acid in an IgE binding site or reacting the allergen with a compound blocking binding to at least one amino acid in an IgE binding site;

(c) screening for IgE binding to the modified allergen using serum or antibodies from a pooled patient population and screening for activation of T cells; and

(d) selecting the modified allergens which have decreased binding to IgE as compared to the unmodified allergen and which activate T cells.

2. The method of claim 1 further comprising screening for binding of the modified allergen for binding to IgG and selecting the modified allergens which have decreased binding to IgE, activate T cells and bind to IgG.

3. The method of claim 1 wherein the modified allergen is mutated in the center of one or more of the IgE binding sites.

4. The method of claim 1 wherein the modified allergen is mutated by substituting a hydrophobic amino acid in the center of one or more of the IgE binding sites with a neutral or hydrophilic amino acid.

5. The method of claim 1 wherein binding of IgE to the modified allergen is blocked by reaction of a compound with at least one amino acid present in an IgE binding site.

6. The method of claim 5 wherein binding of IgE is blocked by reaction of the allergen with an antibody which blocks binding to one or more IgE sites but allows the allergen to still activate T cells.

7. The method of claim 1 wherein the modified allergen is a portion of a protein.

8. The method of claim 1 wherein the modified allergen is formulated with an adjuvant selected from the group consisting of IL 12, IL 16, IL 18, Ifn-γ or immune stimulatory sequences.

9. The method of claim 1 wherein the modified allergen is screened for initiation of a T cell helper 1 response.

10. The method of claim 1 wherein the modified allergen is made in a recombinant host selected from the group consisting of plants, animals, bacteria, yeast, fungi, and insect cells.

11. The method of claim 1 wherein the modified allergen is made in cells using site specific mutation.

12. The method of claim 1 wherein the modified allergen is made from a peanut allergen selected from the group consisting of Ara h 1, Ara h 2, and Ara h 3.

13. The method of claim 1 wherein the modified allergen is based on a protein obtained from a source selected from the group consisting of legumes, milks, grains, eggs, fish, crustaceans, mollusks, insects, molds, dust, grasses, trees, weeds, mammals, birds, and natural latexes.

14. A modified allergen which is less reactive with IgE comprising at least one IgE binding site present in the allergen modified by at least one amino acid change or having at least one amino acid bound by a compound so that the site no longer binds IgE, wherein the modified allergen activates T cells.

15. The modified allergen of claim 14 wherein the modified allergen binds IgG.

16. The modified allergen of claim 14 made by the process of

(a) identifying one or more IgE binding sites in an allergen;

(b) mutating at least one amino acid in an IgE binding site;

(c) screening for IgE binding to the mutated allergen and activation of T cells by the mutated allergen; and

(d) selecting the modified allergens with decreased binding to IgE which activate T cells.

17. The modified allergen of claim 14 wherein the modified allergen is mutated in the center of one or more of the IgE binding sites.

18. The modified allergen of claim 14 wherein the modified allergen is mutated by substituting a hydrophobic amino acid in the center of one or more of the IgE binding sites with a neutral or hydrophilic amino acid.

19. The modified allergen of claim 14 wherein binding of IgE is blocked by reaction of a compound with at least one amino acid present in an IgE binding site.

20. The modified allergen of claim 19 wherein binding of IgE is blocked by reaction of the allergen with an antibody which blocks binding to one or more IgE sites but allows the allergen to still activate T cells.

21. The modified allergen of Claim 20 wherein the modified allergen does not have significantly altered or decreased IgG binding capacity.

22. The modified allergen of claim 14 which initiates a T cell helper 1 response.

23. The modified allergen of claim 14 wherein the allergen is a portion of a protein.

24. The modified allergen of claim 14 wherein the modified allergen is formulated with an adjuvant selected from the group consisting of IL 12, IL 16, IL 18, Ifn-γ and immune stimulatory sequences.

25. The modified allergen of claim 14 wherein the modified allergen is made in a transgenic plant or animal.

26. The modified allergen of claim 14 expressed in a recombinant host selected from the group consisting of plants and animals.

27. The modified allergen of claim 17 expressed in a recombinant host selected from the group consisting of bacteria, yeast, fungi, and insect cells.

28. The modified allergen of claim 14 wherein the modified allergen is based on a protein obtained from a source selected from the group consisting of legumes, milks, grains, eggs, fish, crustaceans, mollusks, insects, molds, dust, grasses, trees, weeds, mammals, birds, and natural latexes.

29. The modified allergen of claim 14 wherein the modified allergen is made from a peanut allergen selected from the group consisting of Ara h 1, Ara h 2, and Ara h 3.

30. A nucleotide molecule encoding a modified allergen which is less reactive with IgE comprising at least one IgE binding site present in the allergen modified by at least one amino acid change so that the site no longer binds IgE, but wherein the modified allergen activates T cells, as defined by any of claims 14, 15, 16, 17, 18, 21 or 22.

31. The molecule of claim 30 in a vector for expression in a recombinant host.

32. A nucleotide molecule for causing a site specific mutation in a gene encoding a protein which yields a modified allergen which is less reactive with IgE comprising at least one IgE binding site present in the allergen modified by at least one amino acid change so that the site no longer binds IgE, but wherein the modified allergen activates T cells.

33. A transgenic plant expressing a modified allergen which is less reactive with IgE comprising at least one IgE binding site present in the allergen modified by at least one amino acid change so that the site no longer binds IgE, but wherein the modified allergen activates T cells, as defined by any of claims 14, 15, 16, 17, 18, 19, 20, 21, 22, or 23.

34. A transgenic animal expressing a modified allergen which is less reactive with IgE comprising at least one IgE binding site present in the allergen modified by at least one amino acid change so that the site no longer binds IgE, but wherein the modified allergen activates T cells, as defined by any of claims 14, 15, 16, 17, 18, 19, 20, 21, 22, or 23.

35. A compound selectively binding to at least one amino acid in an IgE binding site of an allergen, wherein the site no longer binds IgE, but wherein the allergen is able to activate T cells, wherein the compound is obtained using a combinatorial library or combinatorial chemistry and screening for reaction with the allergen to produce bound allergen, followed by testing of the bound allergen for binding to IgE and activation of T cells.

36. A method to treat an individual to reduce the clinical response to an allergen comprising administering to the individual a modified allergen

which is less reactive with IgE comprising at least one IgE binding site present in the allergen modified by at least one amino acid change or having at least one amino acid bound by a compound so that the site no longer binds IgE, wherein the modified allergen activates T cells, as defined by any of claims 14-29, in an amount and for a time sufficient to reduce the allergic reaction to the unmodified allergen.

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INTERNATIONAL SEARCH REPORT

International Application No
PCT/US 99/02031

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12N15/29 A61K39/35 C07K14/415 A01H5/00 A01K67/027

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C12N C07K A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	STANLEY JS ET AL: "Identification and mutational analysis of the immunodominant IgE binding epitopes of the major peanut allergen Ara h 2" ARCHIVES OF BIOCHEMISTRY AND BIOPHYSICS, vol. 342, no. 2, 15 June 1997, pages 244-253, XP002107202 see page 251, left-hand column; figure 5A; table III ---	1-7, 10-14, 16-29

-/--

[X] Further documents are listed in the continuation of box C.

[ ] Patent family members are listed in annex.

° Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
24 June 1999	07/07/1999

Name and mailing address of the ISA	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl, Fax: (+31-70) 340-3016	Cupido, M

1

International Application No
PCT/US 99/02031

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication,where appropriate, of the relevant passages	Relevant to claim No.
X	BURKS AW ET AL: "Mapping and mutational analysis of the IgE-binding epitopes on Ara h 1, a legume vicilin protein and a major allergen in peanut hypersensitivity " EUROPEAN JOURNAL OF BIOCHEMISTRY, vol. 245, no. 2, April 1997, pages 334-339, XP002107203 see page 336 - page 339	1-7, 10-14, 16-29
A	BURKS AW ET AL: "Recombinant peanut allergen Ara h 1 expression and IgE binding in patients with peanut hypersensitivity" THE JOURNAL OF CLINICAL INVESTIGATION, vol. 96, no. 4, October 1995, pages 1715-1721, XP002107204 see page 1717 to 1720, especially page 1720 first paragraph	1-34

1

INTERNATIONAL SEARCH REPORT	International application No. PCT/US 99/02031

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. [X] Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
Remark: Although claim 36 is directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.

2. [X] Claims Nos.: 35
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
The subject-matter of the claim is unsufficiently characterised to allow a meaningful search

3. [ ] Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. [ ] As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.

2. [ ] As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. [ ] As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:

4. [ ] No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

[ ] The additional search fees were accompanied by the applicant's protest.

[ ] No protest accompanied the payment of additional search fees.